



BioFrontiers Institute
UNIVERSITY OF COLORADO **BOULDER**



User Training: The Foundation of a Successful Core

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RRID: SCR_018302

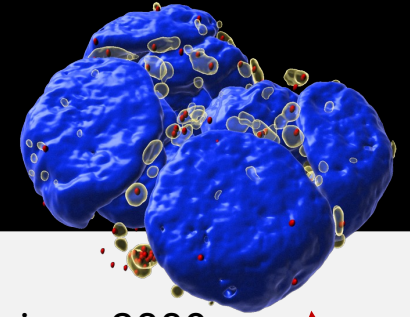
Who am I and what experience do I have?



Dr. Joe Dragavon
Director
Advanced Light Microscopy Core
BioFrontiers Institute

- Worked microscopy Cores since 2009
- Became Director of the ALMC in 2014

Who am I and what experience do I have?

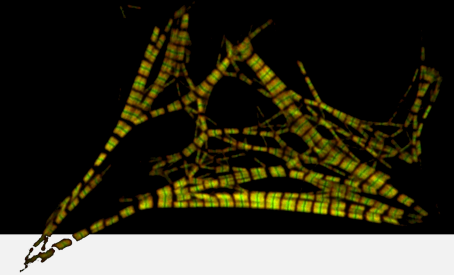


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If you get bored,
pretty images can
be found here

Who am I and what experience do I have?



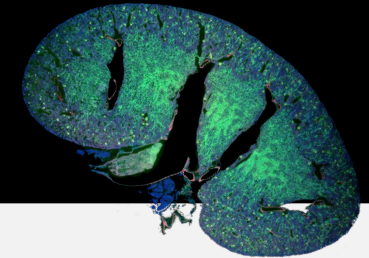
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ALMC 10-year Stats

	3-Year Average	10-Year Total
Unique Users	175	729
Unique Labs	59	126
Hours of Use	12,341	126,854
Trainings	123.7	A lot (825)

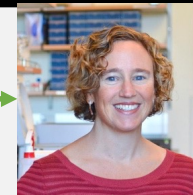
BFALMC Structure (2023)



Dr. Joe Dragavon
Director
(2014)



Prof. Amy Palmer
Faculty Advisor



Dr. Jian Tay
Image Analysis
Specialist
(2017)



- Day-to-day operations
- Facility management
- Beckman Center for Light Sheet Microscopy Oversight
- QOI Co-Lecturer
- Instrument maintenance and support
- User trainings and support
- Commercial Image Analysis support
- Grant writing
- Strategy development
- Building instrument support

Dr. Evolene Premillieu
Beckman Center
Light Sheet Specialist
(2023)



Responsibilities

- Custom Image Analysis (MATLAB)
- QOI Co-Lecturer
- General analysis guidance and facility support
- Web Support

- Bruker TruLive 3D
- Single Objective Light Sheet
- Web Support

Guidance
Supervision + Responsibilities

User Training – Part of the Core Circle of Life

More Users = More Trainings



A robust training program is the **foundation for my Core**

As your reputation grows, **more PIs** will want to work with you. This will result in **additional and consistent Users**.

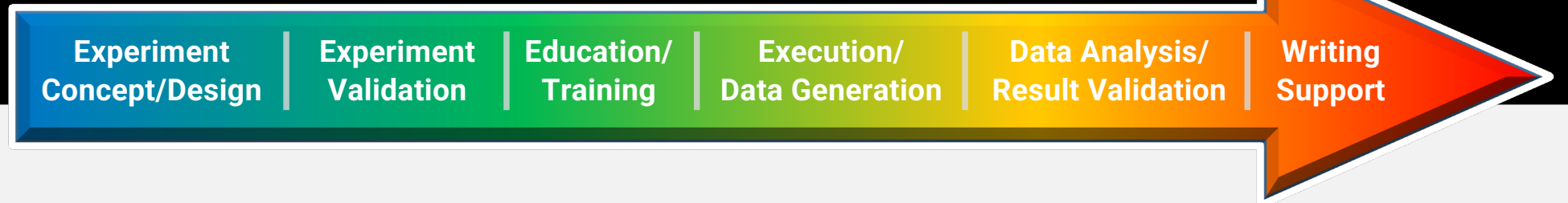
Use the training to form a **professional relationship** to make your more integral to their success.

If your Users trust you, so will their PI. Over time, you become a valuable resource with a solid reputation.

Assist your Users to facilitate Research Success. **Your Users will rely on you for this.**

Relationships are built on trust: **Helping the User become successful will mean they will trust you feedback more** (and they will tell others!).

Onboarding and Support Pipeline



1. Initial meeting for Experiment Design

- Concepts/goals discussed
- Appropriate experiment(s) designed

2. Validation Experiments

- Controls are tested
- Acquisition workflow validated

3. Education and Training

- Microscopy theory
- Train to autonomy
- 3 - 6 hours each

4. Execution and Data Acquisition

- Troubleshooting
- Experiment refinement

5. Image and Data Analysis

- Statistics
- Automated analysis
- Robustness/efficiency

6. Writing Support

- Grant applications
- Publications

Onboarding and Support Pipeline

Experiment
Concept/Design

Experiment
Validation

Education/
Training

Execution/
Data Generation

Data Analysis/
Result Validation

Writing
Support

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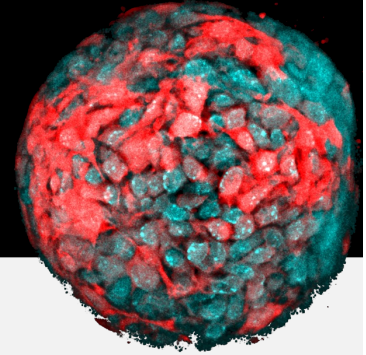
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- Statistics
- Automated analysis
- Robustness/efficiency

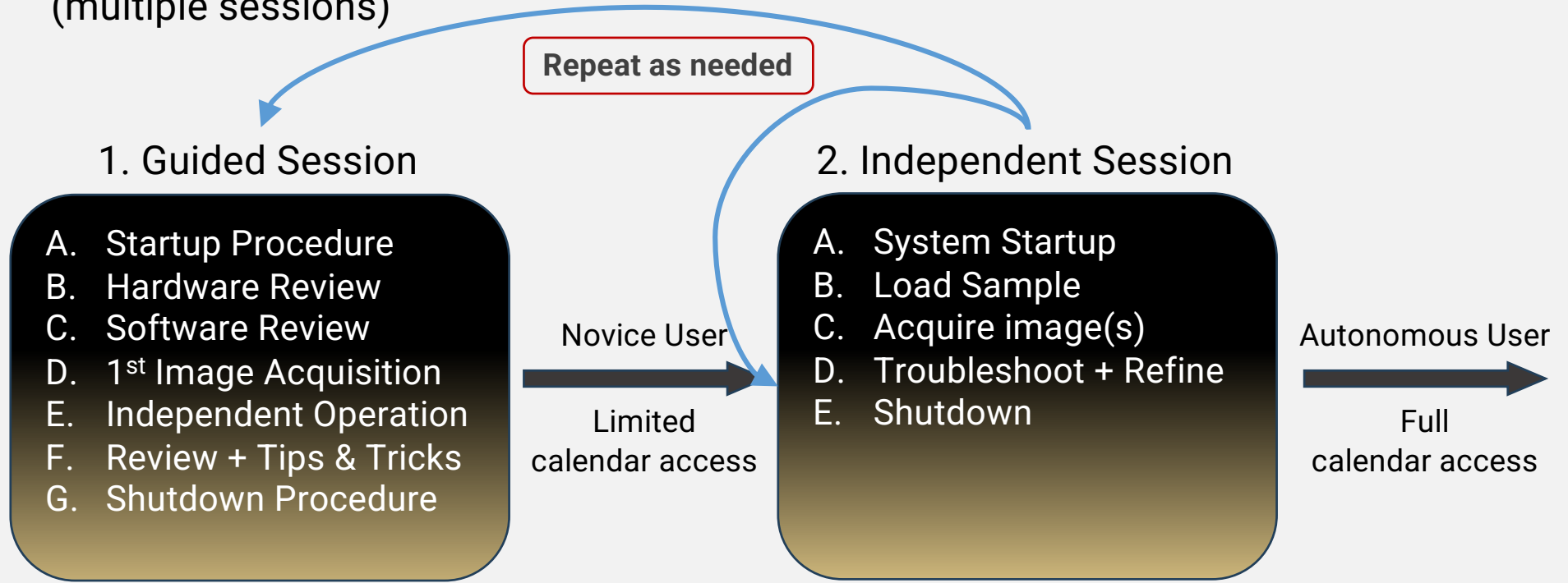
6. Writing Support

- Grant applications
- Publications

3. Education and (long, but slow) Training

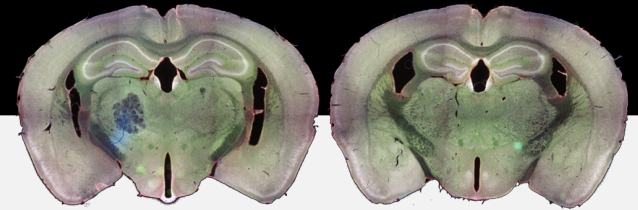


My trainings take 3 – 6 hours (or more), depending on the system.
(multiple sessions)



3. Education and (long, but slow) Training

Part 1: Guided Session



A. Startup Procedure

- This should be posted near the instrument, and switches labeled
- The User should power on the system

B. Hardware review

- What are all the boxes?
- How to load a sample
- How to navigate using the controller

1. Guided Session

- A. Startup Procedure
- B. Hardware Review
- C. Software Review
- D. 1st Image Acquisition
- E. Independent Operation
- F. Review + Tips & Tricks
- G. Shutdown Procedure

Experiment
Concept/Design

Experiment
Validation

Education/
Training

Execution/
Data Generation

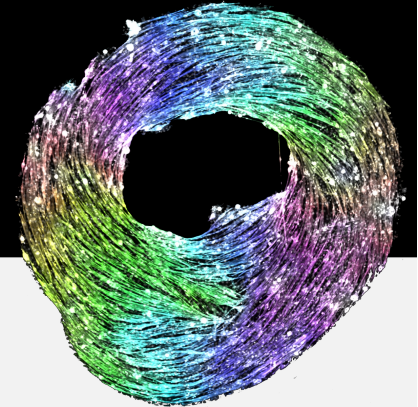
Data Analysis/
Result Validation

Writing
Support



3. Education and (long, but slow) Training

Part 1: Guided Session



C. Step-by-step software review

- Go over all the panels of the software BEFORE pushing a button
- At certain steps, slow down and explain the finer details/importance
 - What is pixel binning and why is it important?
 - What is saturation?
 - What is Nyquist sampling?

D. User-led/instructor-guided sample collection

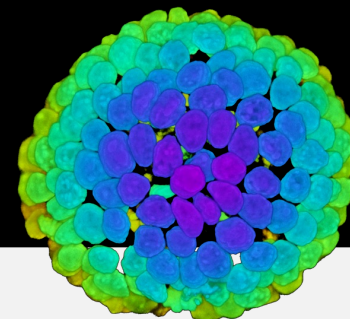
- Have the User load your standard sample
 - I use the same sample every time (I know that it will work)
 - Surprises during a training create confusion and doubt
- Have the User control the mouse and click the buttons
- Tell them what will happen BEFORE they click something

1. Guided Session

- A. Startup Procedure
- B. Hardware Review
- C. **Software Review**
- D. **1st Image Acquisition**
- E. Independent Operation
- F. Review + Tips & Tricks
- G. Shutdown Procedure

3. Education and (long, but slow) Training

Part 1: Guided Session



E. Independent operation

- Turn the software off and remove the sample
- On their own, have the User load the sample and collect a standard set of images
 - E.g. Tiled images, multicolor images, Nyquist
 - I want the User to feel quite independent
 - Note: I leave the room during this part for at least 10 minutes

F. Operation Review and Tips and Tricks

- Do the images look correct?
 - As I know the sample, I know what correct images should look like
- Provide tips and tricks to improve image quality/performance
- Repeat independent acquisition if necessary

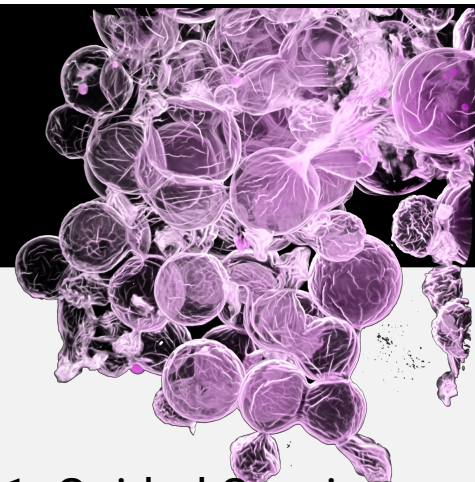
1. Guided Session

- A. Startup Procedure
- B. Hardware Review
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3. Education and (long, but slow) Training

Part 1: Guided Session

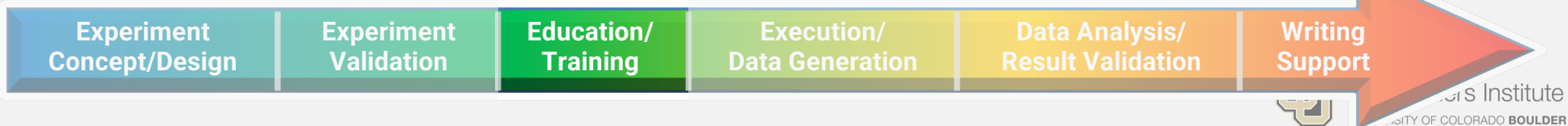


G. Review Shutdown Procedure

- This should also be posted
- Include how they should contact you if there is a problem!
 - **I include my personal cell phone number**
- Remind the User you are there to help
 - Their success is my success
 - **When my Users are working, I am on call**

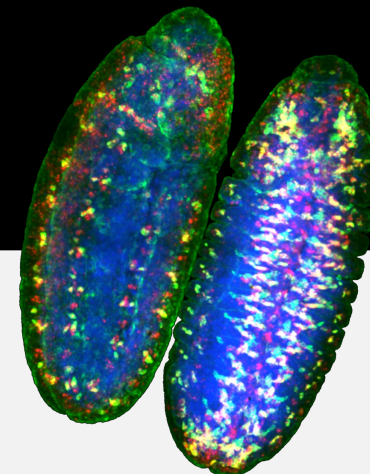
1. Guided Session

- A. Startup Procedure
- B. Hardware Review
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3. Education and (long, but slow) Training

Part 2: Independent Session

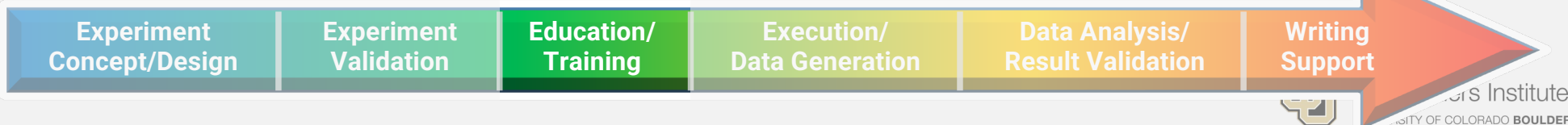


A. Coordinate with the User for the Independent Session

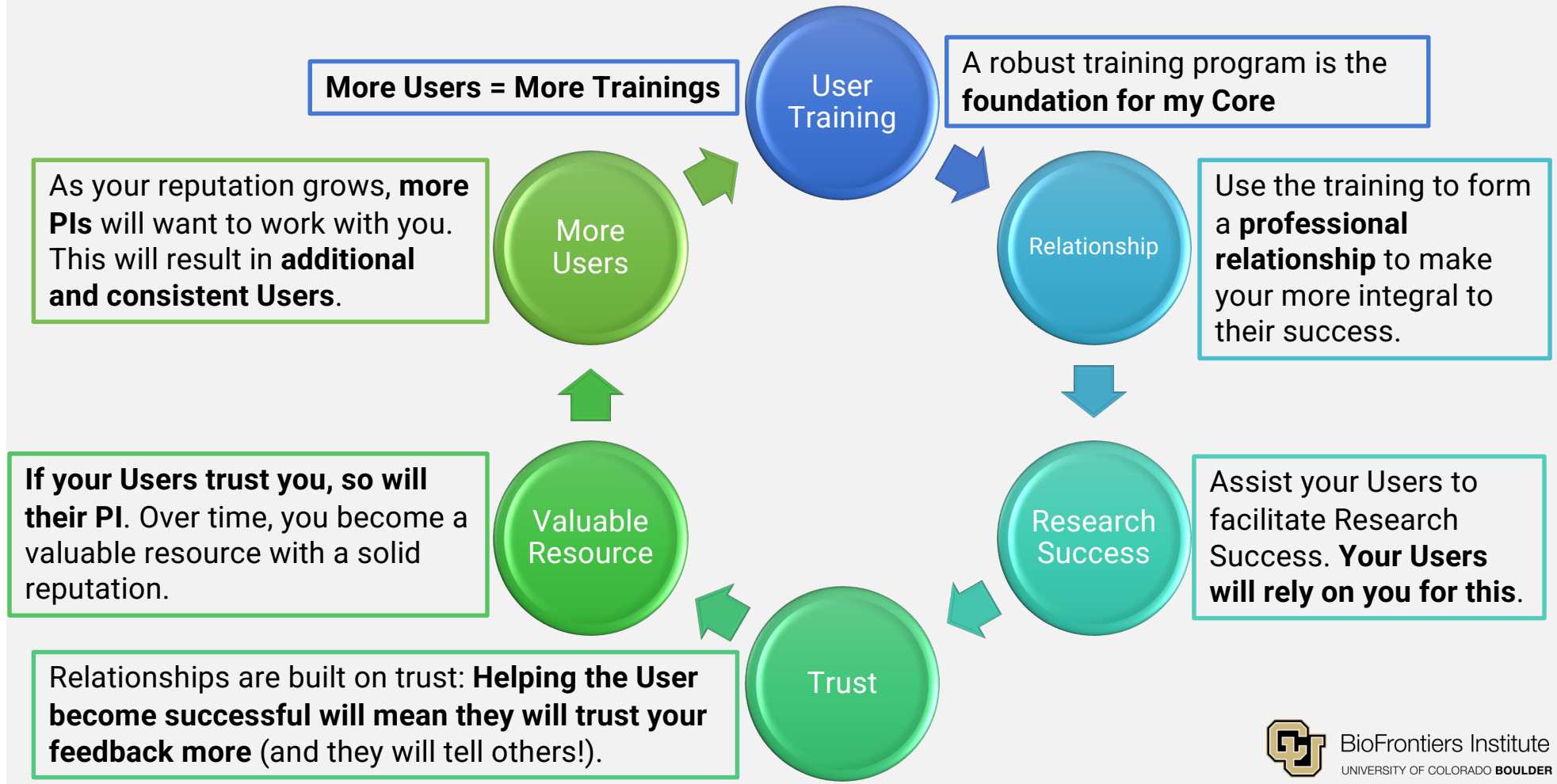
- I want to be available to help, but not hovering in the room
- Check in multiple times throughout the session
- The User needs to gain confidence in what they are doing
 - **I have other things to do**
- Remind the User you are there to help

2. Independent Session

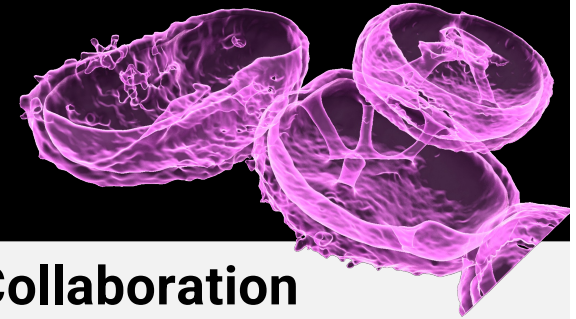
- A. System Startup
- B. Load Sample
- C. Acquire image(s)
- D. Troubleshoot + Refine
- E. Shutdown



User Training – Part of the Core Circle of Life

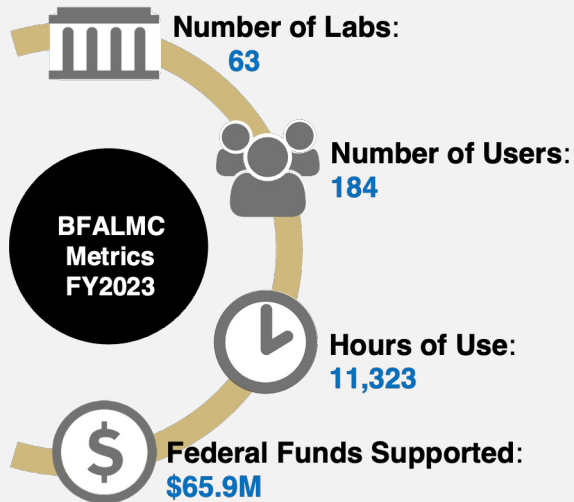


Facility Impact



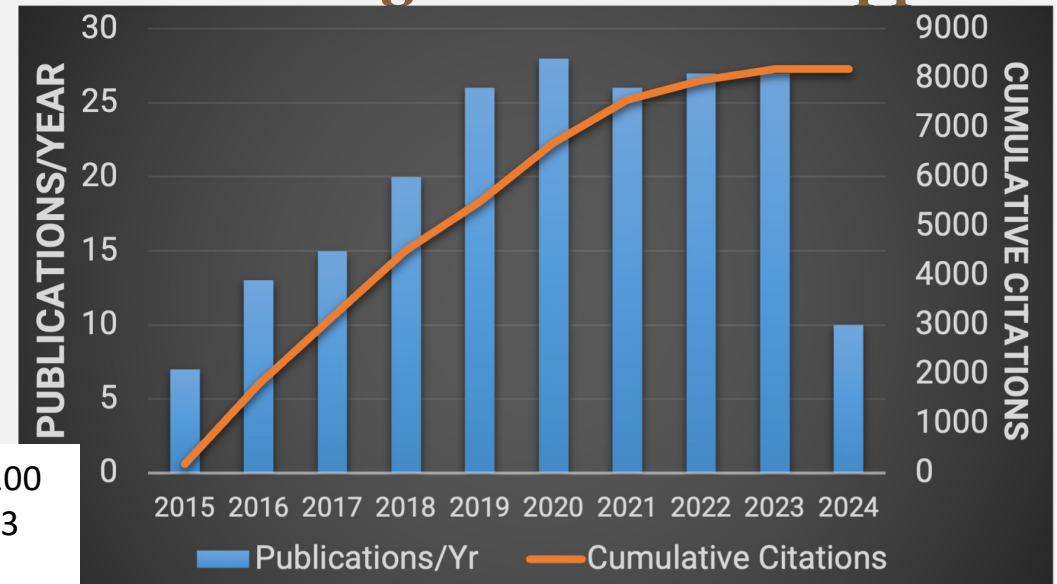
We are more than a Service Center, we are a **Center for Collaboration**

2023 University-Wide Impact



Publications: 200
Citations: 8,223
H-Index: 43

Number of CU publications resulting from ALMC support



Acknowledgements



BioFrontiers Institute
UNIVERSITY OF COLORADO BOULDER

BioFrontiers and CU Boulder

All Faculty and Staff
ALMC Members

- Jian Tay
- Evolene Premillieu
- Amy Palmer

All my Users+PIs+Dept Chairs
Research and Innovation Office

- Karen Regan
- Massimo Ruzzene
- Alicia Aldeman



External Funding



Thank you for listening



ABRF Colleagues

- Ashlyn Montgomery
- Andy Chitty
- Claudius Mundoma
- Rich Cole
- Mark Sanders
- Sheena Mische
- Susan Weintraub
- LMRG, EdComm, 2024 PC

Probing Questions

A. Pre-training content?

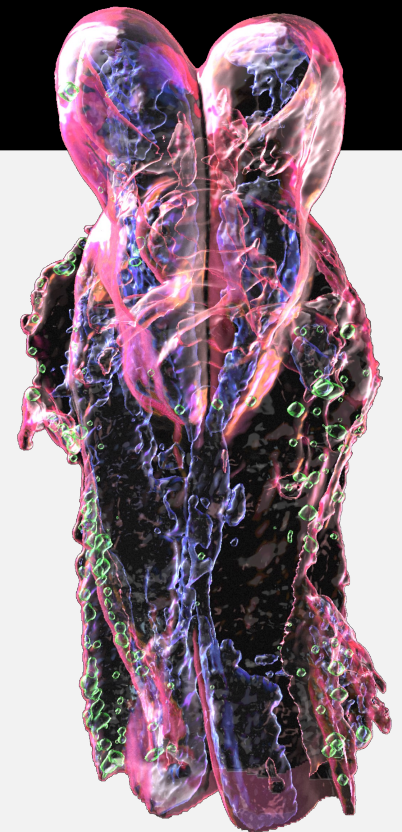
- Presentations/videos/websites
- Pre-training quiz?

B. Post-training quiz/follow-up?

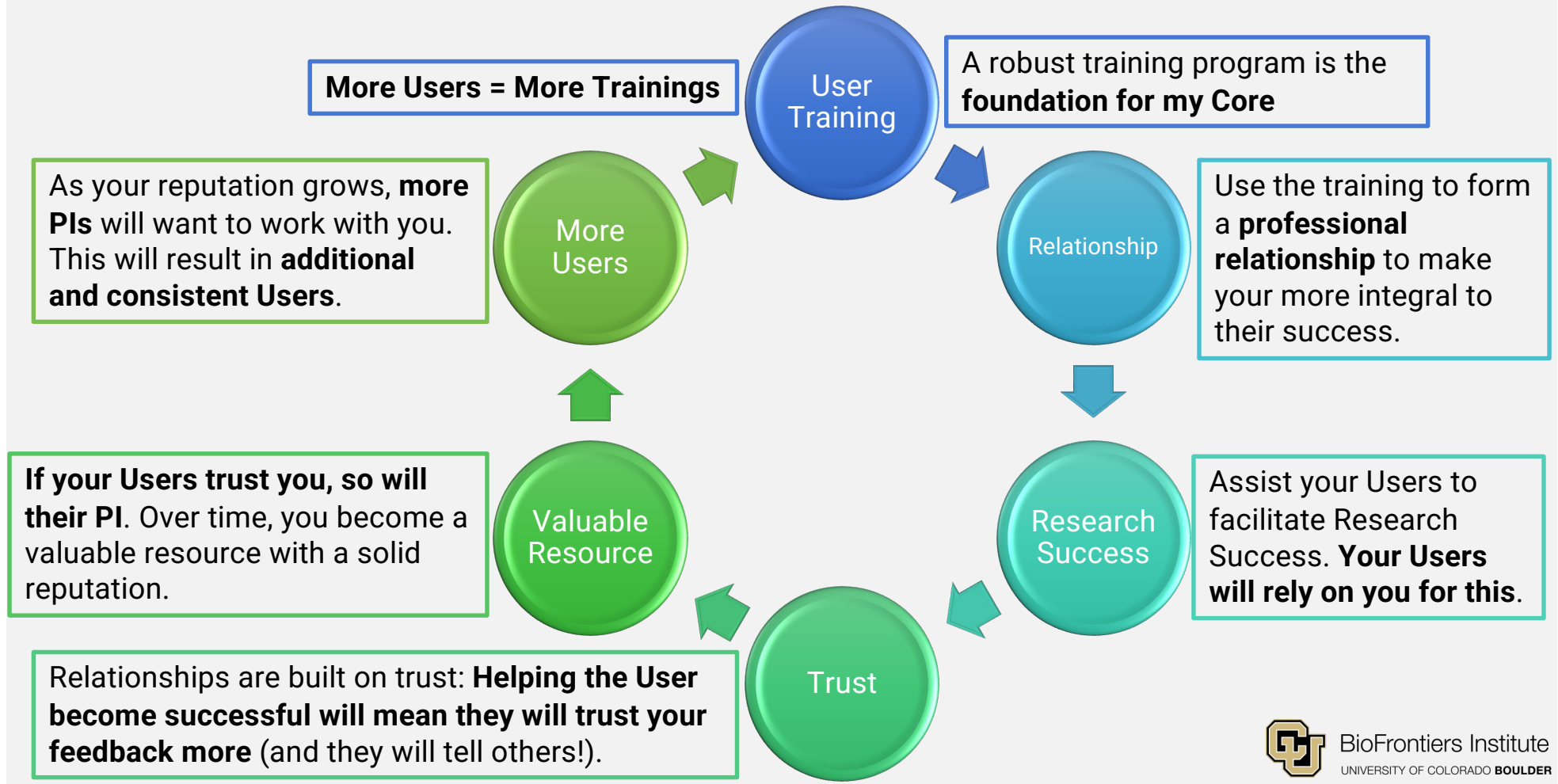
- How do you evaluate if a User is ready?

C. Start with Standard Samples or User Samples

- Depends on the technology
- Many Users don't know how to prepare samples

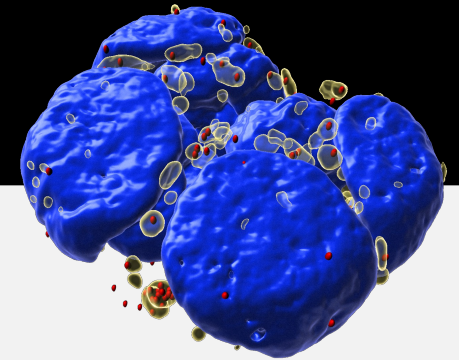


User Training – Part of the Core Flywheel Effect



Bonus Slides

1. Initial Meeting for Research Discussion



A. Get to know the User

- Face-to-face meeting
- Gauge their understanding of the research and technology

B. Experimental design review

- Concepts/goals discussed
- Appropriate experiments/controls designed

C. Set trial and training day/time

- Find a time that works for us (and the instrument)
- Preview what is involved in the training
- Advise the User on what to bring

Experiment
Concept/Design

Experiment
Validation

Education/
Training

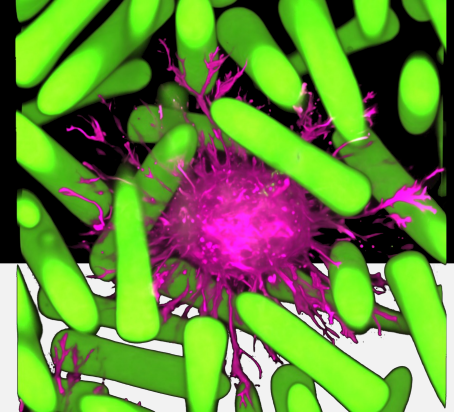
Execution/
Data Generation

Data Analysis/
Result Validation

Writing
Support



2. Validation experiments



A. Start with the controls

- Demonstrating the instrument on the extremes builds confidence
- Provides some immediate data

B. Talk while pushing the buttons

- Introduces the User to the system, removing some of the mystique
- If possible, allow the User to collect an image or two

C. Get to know the User more

- Demonstrate that you are knowledgeable, but approachable

Experiment
Concept/Design

Experiment
Validation

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Data Generation

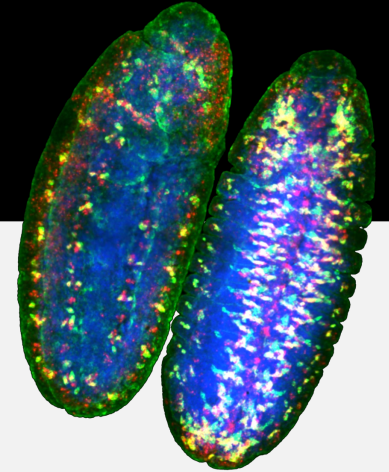
Data Analysis/
Result Validation

Writing
Support



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4. Execution/Data Generation



A. Independent sessions

- Review the calendar
 - Keep an eye on your newer/less confident Users
- Walk through the facility regularly
 - Users can seem reticent to reach out if they are uncertain
 - So, watch them work and help out in real time!
- Promote the acknowledgement of the Core (maybe mention co-authorship...)

Experiment
Concept/Design

Experiment
Validation

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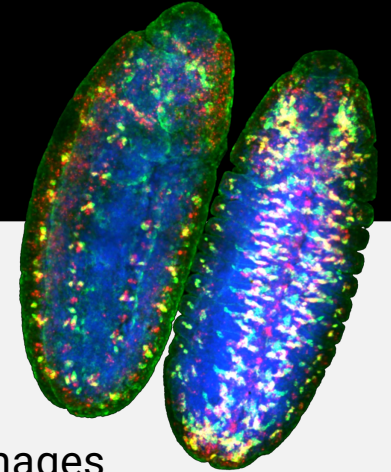
Writing
Support



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5. Data Analysis/Result Validation

6. Writing Support

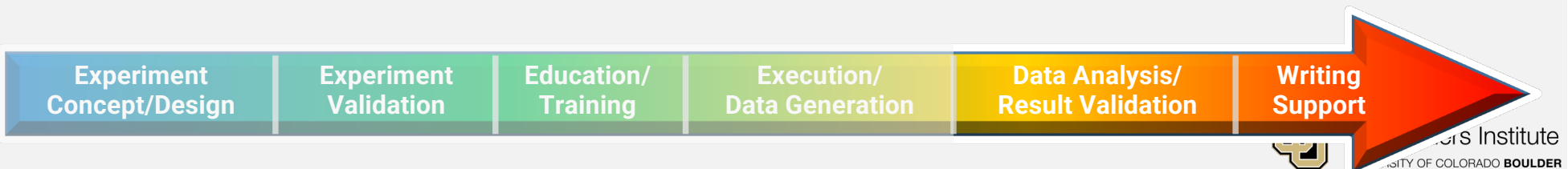


A. Users need help analyzing their data

- Provide guidance/point them towards relevant resources
 - Many of us don't have enough bandwidth to do a deep dive here
- Talk about proper lab notebook keeping and to regularly analyze their images

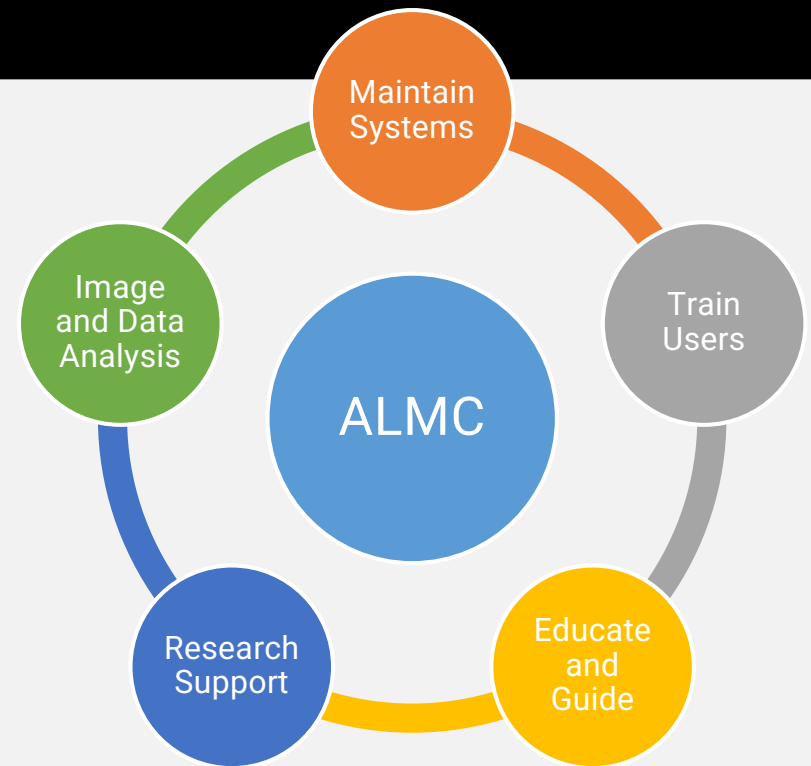
B. Offer to review the relevant sections of their papers

- Ensure proper details are in the methods
- Make sure you are correctly acknowledged (your RRID should be included)



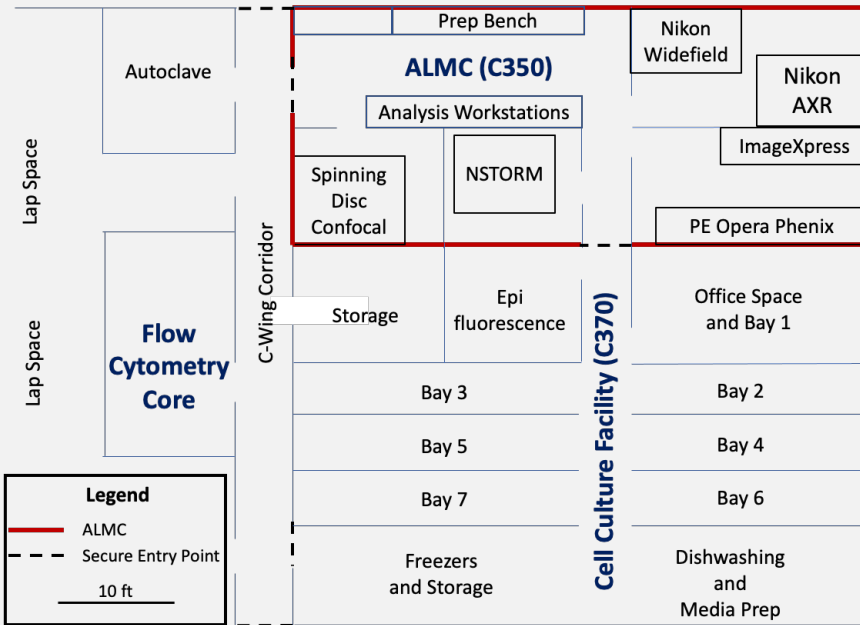
Our Mission:

1. Advance the available imaging and analysis capabilities to meet the needs of our current and future investigators
2. Promote microscopy and data analysis education to increase rigor and impact
3. Facilitate the success of all that may need our resources by enabling access and by providing expert guidance and assistance
4. Increase the efficiency of our research by reducing duplication and maximizing space utilization



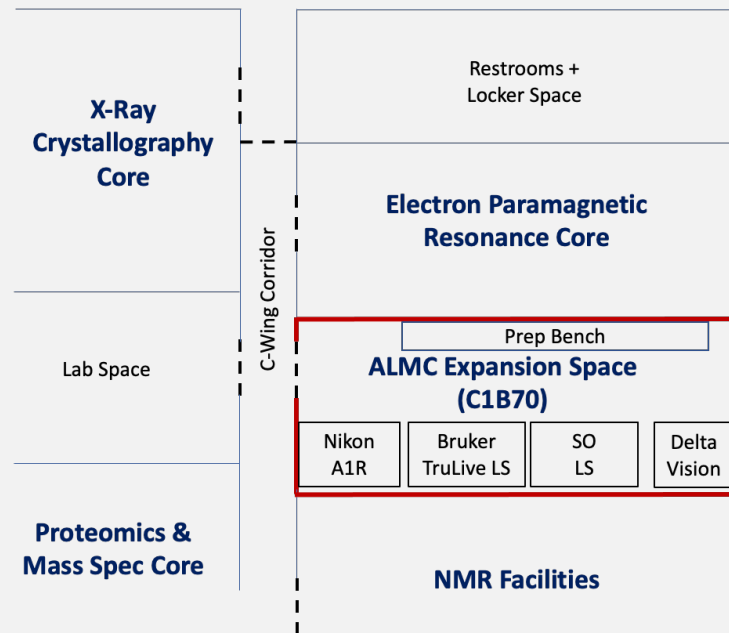
ALMC Location

JSCBB 3rd Floor, C-wing Main Corridor



Total Space: 1300 sq ft

JSCBB Basement, C-wing Main Corridor



+ IVIS Lumina in the vivarium

Research

Management

Administration

Education



Research & Innovation Office
UNIVERSITY OF COLORADO BOULDER

CU Boulder

Shared Instrumentation Network

Find the
TECHNOLOGY, INSTRUMENTATION & EXPERTISE
to ensure your investigative success!

Technical Areas

- » Light + Electron Microscopy & Flow Cytometry
- » NMR & Mass Spectrometry
- » Materials Characterization & Nanofabrication
- » Shared Instrumentation & Resource Centers

Discover the Resources



www.colorado.edu/sharedinstrumentation/



Shared Instrumentation Network



Microscopy and Imaging

- Light Microscopy Core Facility
- BioFrontiers Advanced Light Microscopy Core
- Biochemistry Krios Electron Microscopy
- Boulder Electron Microscopy Services Core Facility
- Flow Cytometry Shared Core
- Intermountain Neuroimaging Consortium

Shared Instrumentation + Resource Centers

- BioCore: Shared Instrumentation
- Shared Instruments Pool in Biochemistry
- Biochemistry Cell Culture Facility
- Stem Cell Research & Technology Resource Center
- Green Labs Shared Ultra-Low Temperature Freezers
- CU Research Computing

NMR and Mass Spectrometry

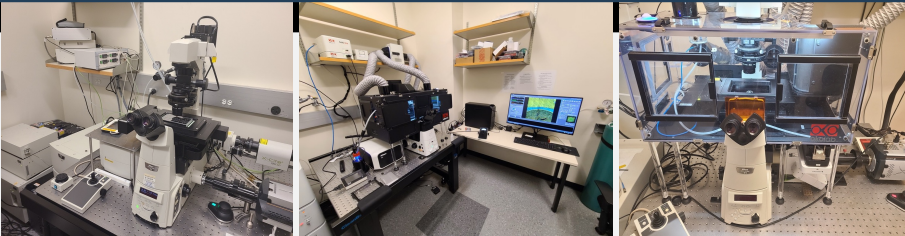
- Biochemistry Nuclear Magnetic Resonance Facility
- Central Analytical Laboratory and Mass Spectrometry Facility
- Nuclear Magnetic Resonance Facility
- Stable Isotope Mass Spectrometry Facility

Materials + Biological Characterization and Nanofabrication

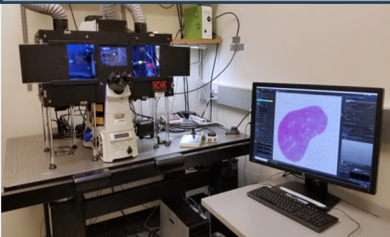
- Colorado Shared Instrumentation in Nanofabrication and Characterization
- CU Electron Microprobe Laboratory
- CU Facility for Electron Microscopy of Materials
- Biomolecular X-ray Crystallography Core
- Materials Research X-ray Diffraction Facility
- Materials Instrumentation and Multimodal Imaging Core
- Polymeric and Optical Materials Characterization Shared Facility
- W. M. Keck Optical Metrology Lab and JILA Micro/Nanofabrication Facility
- Raman Microspectroscopy Laboratory

Our 10 imaging systems

Laser Scanning and Spinning Disc Confocal



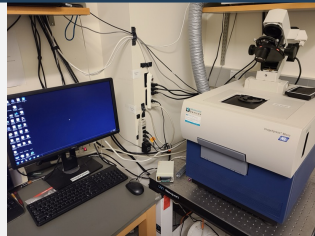
Widefield



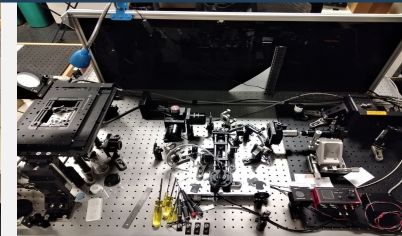
TIRF and Super Res



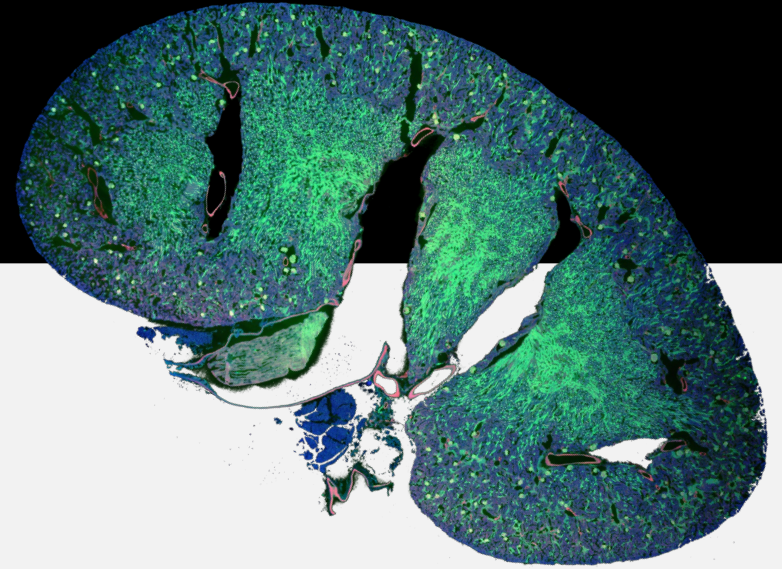
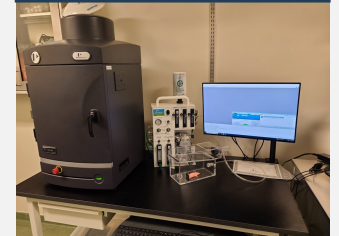
High Throughput/High Content



Light Sheet



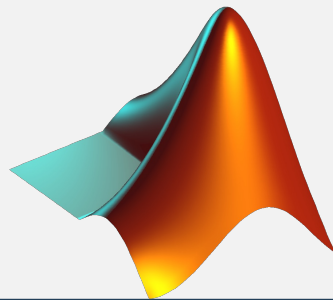
Whole Animal



Our Analysis Tools



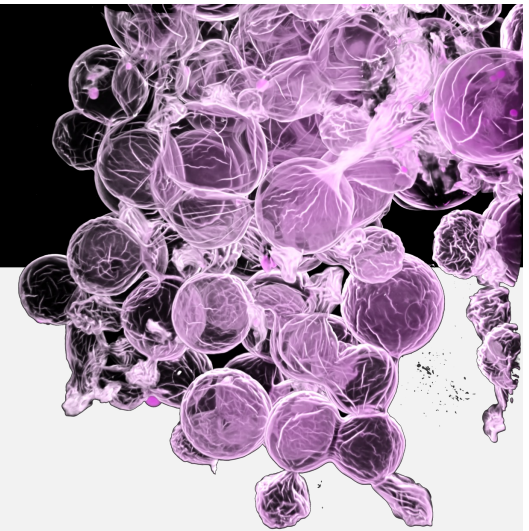
ImageJ/Fiji



MATLAB



Harmony



Imaris



NIS Elements



Cell Profiler